In-vitro Mobilizierung von Kalzium-, Eisenund Aluminium- Phosphat durch Rhizosphärenbakterien der Afrikanischen Ölpalme

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African oil palm Elaeis guineensis

- one of the most important oil crops in the world



juvenil



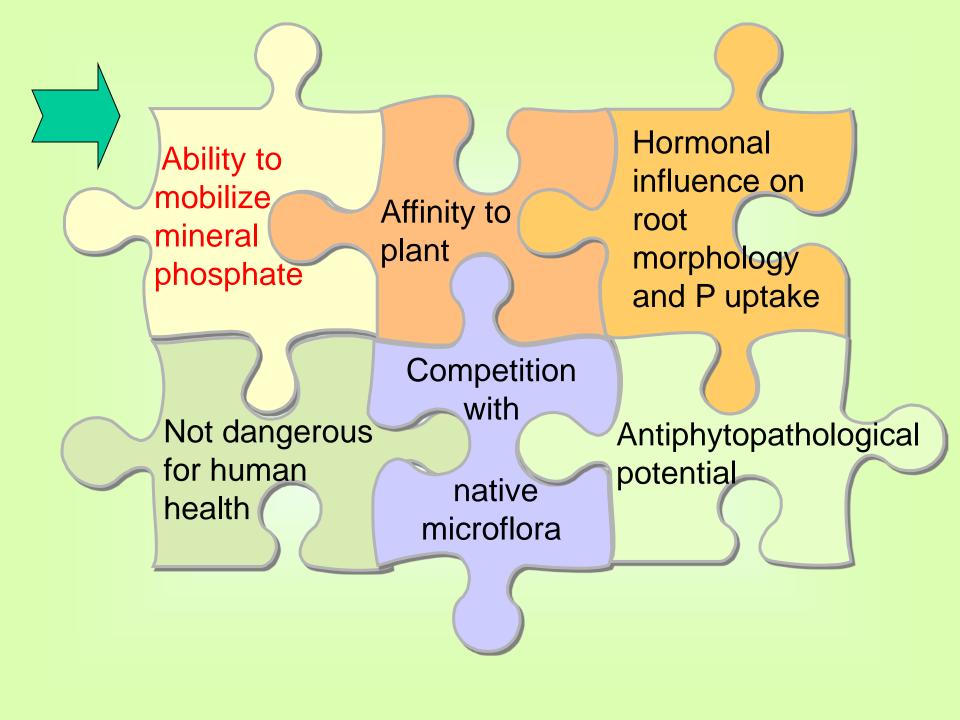
adult

Aim of investigation:

Looking for rhizosphere microbes, which are able to mobilize Ca, Al and Fe from soil for oil palm nutrition

Basis for bio-fertilizer?

Which properties are included in potentially biofertilizer strains?



Question (2)

Why is ability for phosphate mobilisation important for oil palm?

Problem

- phosphate supply is one of the most important yieldlimiting factors in many tropical and subtropical soils
- high reactivity with Ca oder Fe / Al
- mineral fertilizers hardly available
- mycorrhiza needs 1-4 months to be effective, expensive inoculates

Therefore



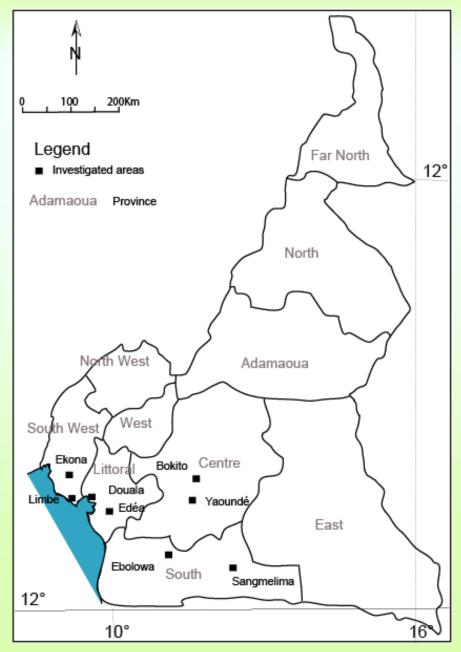
Examination of the occurrence of phosphatemobilising bacteria in the rhizosphere of oil palm

How was determined the phosphate mobilization of mineral phosphate by bacteria?

Step 1

 Isolation of bacterial strains from rhizosphere soil (187) and root fragments (90); repeated inoculation for purity and stability

- Fig. Distribution of the investigated areas in humid-forest zones of Cameroon. These areas are located in two main agro-ecological zones of Cameroon:
 - a) the humid forest zone with one rainy season and that includes provinces of Littoral and South west
- b) the humid forest zone with two rainy seasons that consists of Centre, South and East provinces. These areas represent the main oil palm plantations.

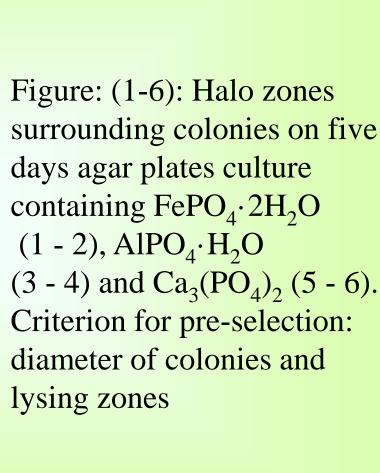


Step 2

•Qualitative test on ability to solubilize Ca₃(PO₄)₂, FePO₄ und AlPO₄ using agarplates with dye(bromo cresol green); evaluation of halo (lysing) zones



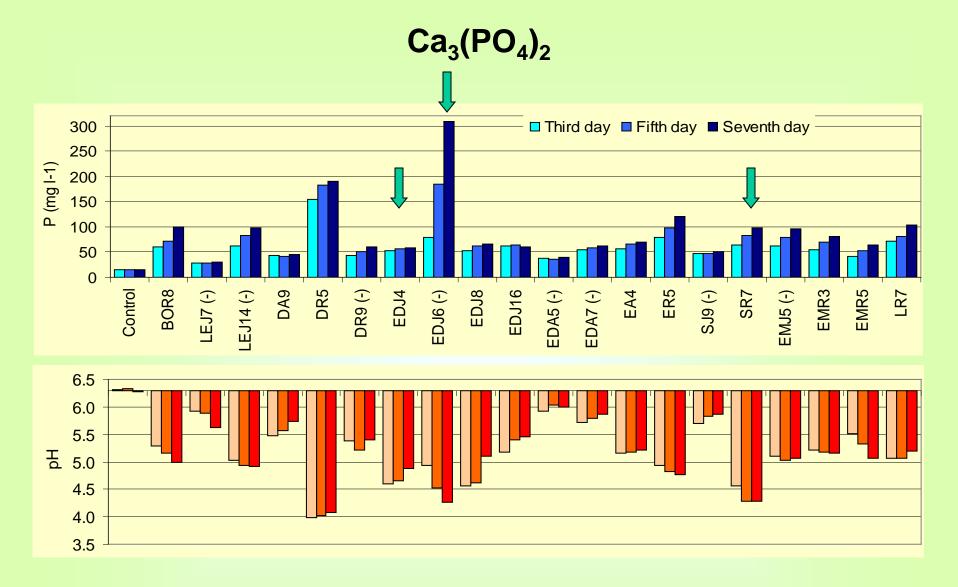




Step 3

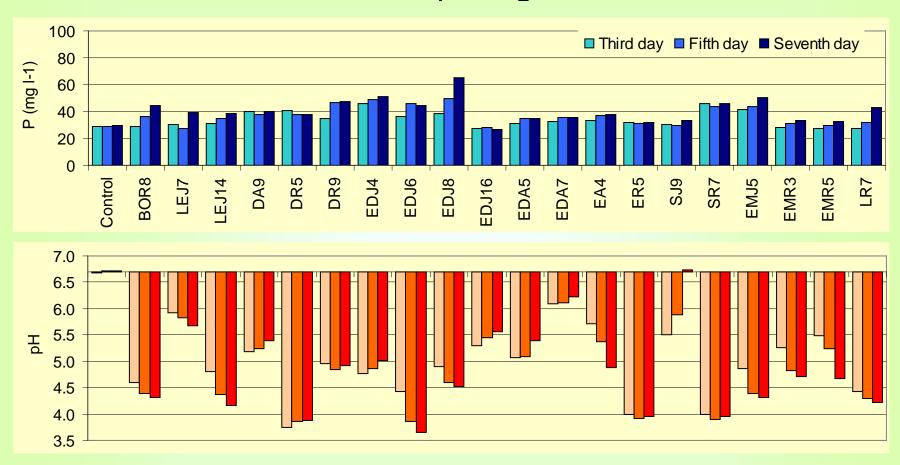
•Quantitative determination of P release fom Ca₃(PO₄)₂, FePO₄ or AIPO₄ in liquid culture. Soluble P was determined by colometry

20 selected strains are proved (for comparison also strains without halo zones)



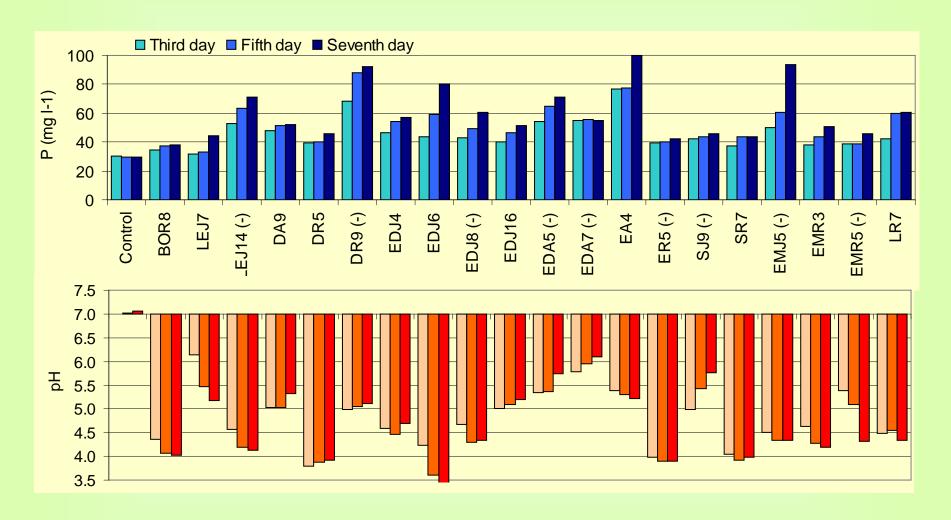
 P releasing was correlated –as a rule –with ph decrease, but there were differences between both properties (see arrows)

FePO₄ · 2 H₂O



Only some strains mobilized P, despite strong pH decreases

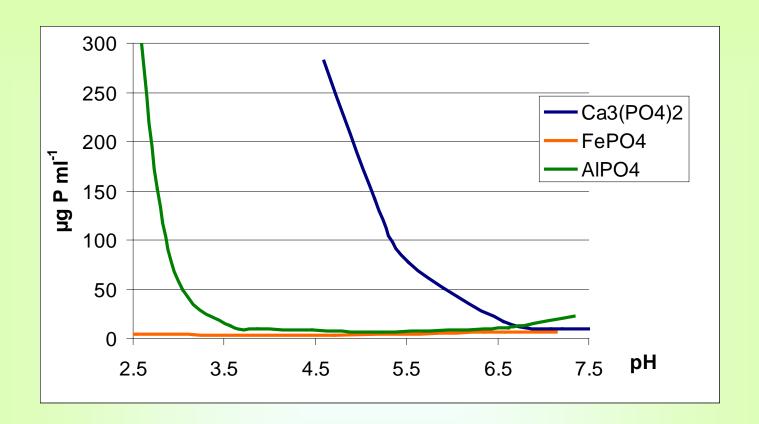
AIPO₄ · H₂O



All strains released P, but no correlation with pH decrease

How does the pH affect the solubility of these phosphates?

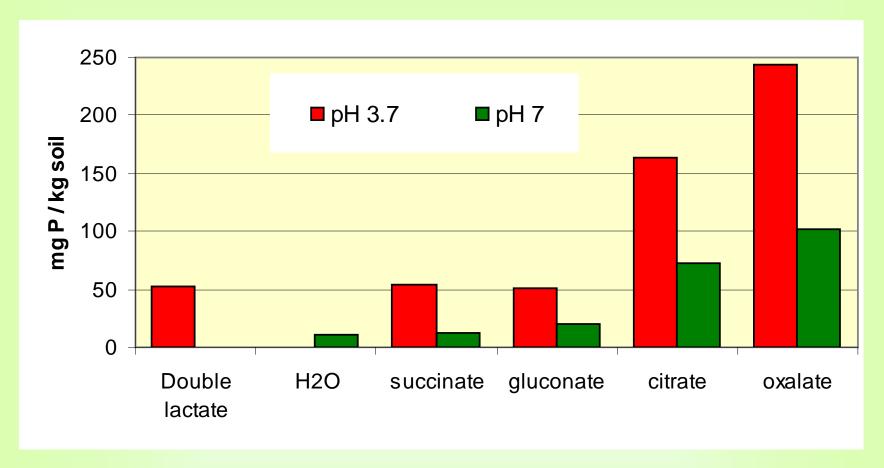
To prove this question, the 3 phosphates were extracted with HCL and NaOH of different concentrations



- CaP solubility increases exponentially with decreasing pH (can explain P release in vitro partly)
- AIP only solubilised at pH levels lower than 3.5 (impossible in bacterial cultures and not useful in plant rhizosphere; could explain P release with some fungal strains)
- Acidification no explanation for FeP solubilisation :not soluble at all pH

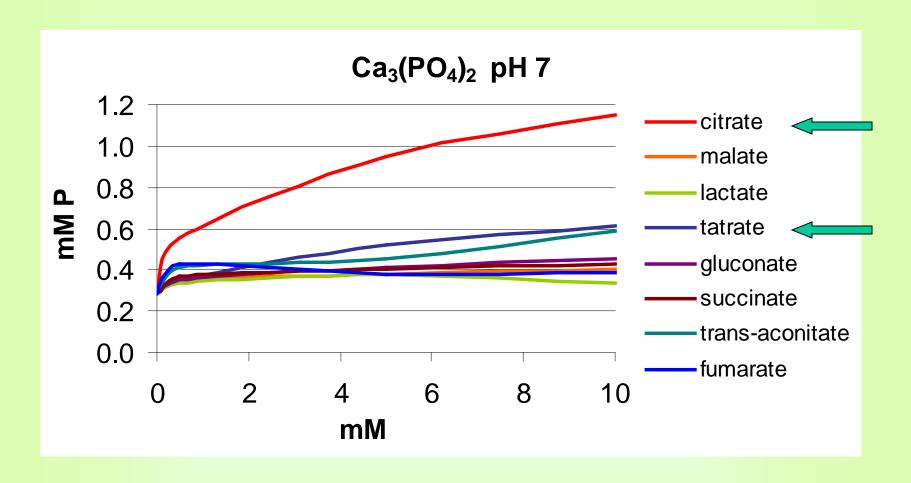
Are carboxylic acids (partly) responsible for P solubilisation? Which carboxylic acids solubilise P under natural soil-pH conditions and in vitro?

Soil conditions

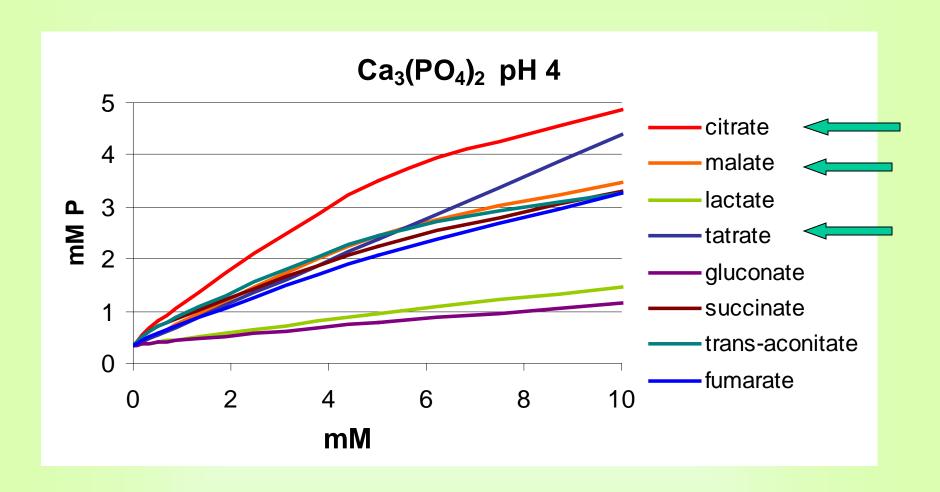


(90 min extraction of 5 g soil with 250 ml solution containing 5 g l⁻¹ of different substances, average from 10 different soils) Carboxylic acids were able to more P than double lactate, esp. at pH 3.7

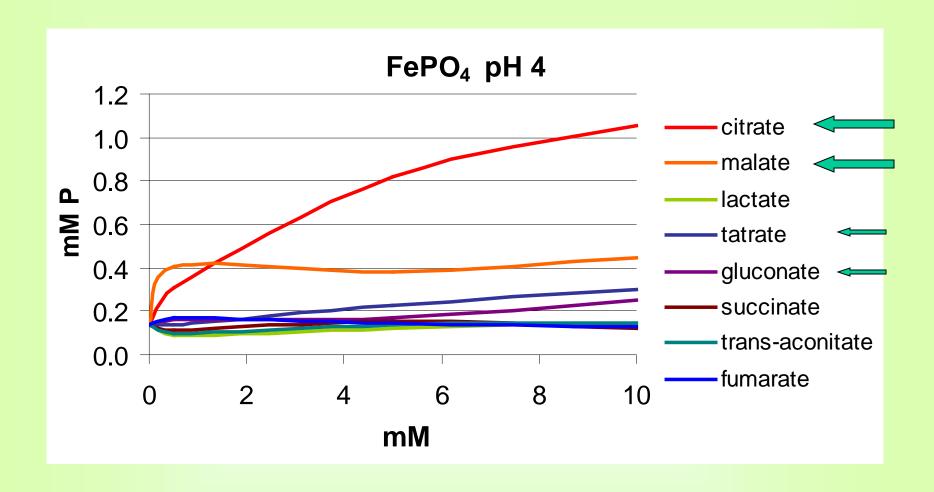
Next step therefore:
In vitro examination of different
carboxylic acids for solubilization of
Ca-, Fe- and Al phosphates



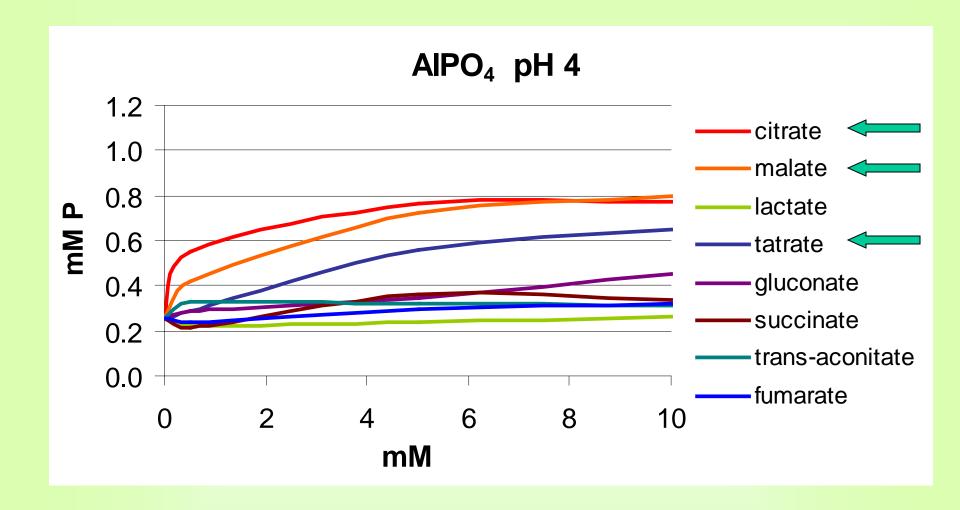
At pH 7: Only citrate increased CaP solubility remarkably



At pH 4 (like in bacterial cultures) some acids increase CaP releasing and complete pH effect. **But**: sparingly soluble CaP occurs in neutral or alkaline soils



Iron phosphate was mobilised by citrate and malate, in a smaller extend by tartrate and gluconate

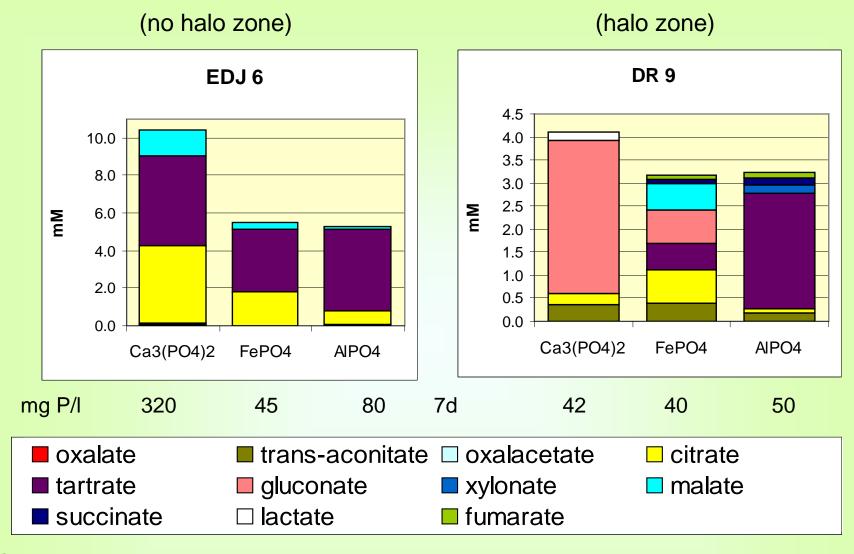


AIP by citrate, malate, tartrate > gluconate.

Acidification no explaination for P solubisation

➤ Which carboxylic acids were produced by bacterial strains and influenced the P releasing?

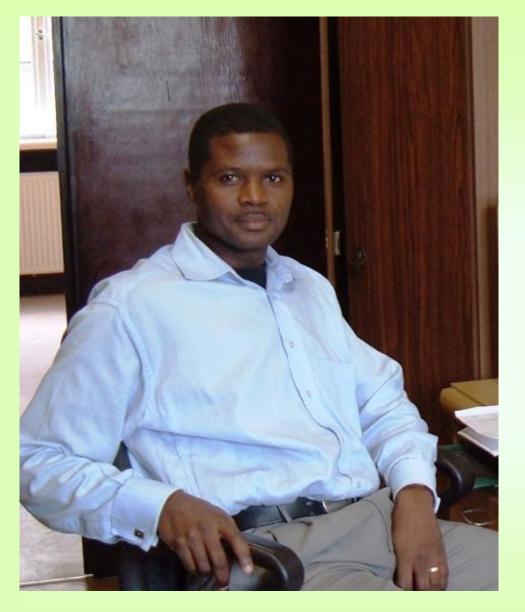
This seems important in case of Al or Fe phosphates (no or low phinfluence of pH value)



Strains produced lot of carboxylic acids and mobilized P. EDJ 6 = most efficient CaP mobilizer: High citrate production and missing halo zone (precipitation of Ca citrate in agar plate test? Limit for pre-selection at agarplates. Substitution by citrate test?). Citrate, tartrate, gluconate most effective.

Conclusions

- P-solubilizing bacteria occur in the rhizosphere of *Elaeis* guineensis; role of microbial P mobilization can be assumed
- Ca₃(PO₄)₂-solubilization in vitro by acidification and different carboxylic acids; at pH 7 (natural soil conditions) mainly by citrate
- FePO₄ and AIPO₄ solubilization by citrate, malate, tartrate, (gluconate)
- Citrate can prevent halo zones by precipitation of Ca-citrate!
- Screening for citrate production would improve the selection of potential P-mobilizing microorganisms



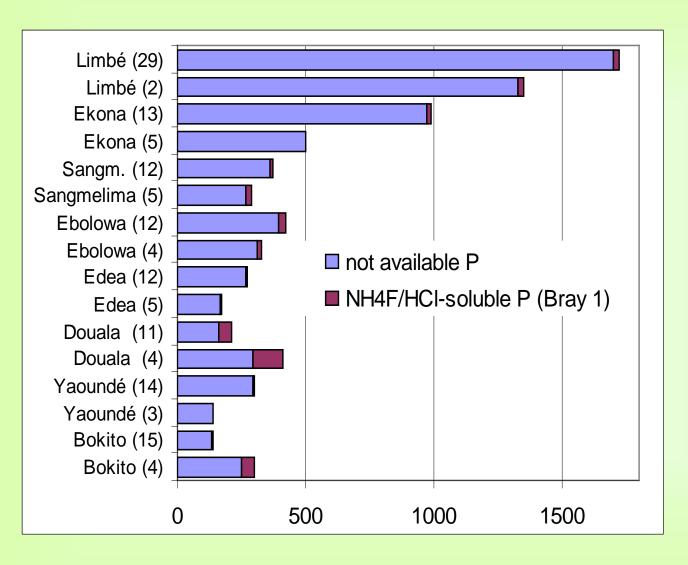
Thank you to all the colleagues

First of all:

Dr.Henri Fankem, Cameroon

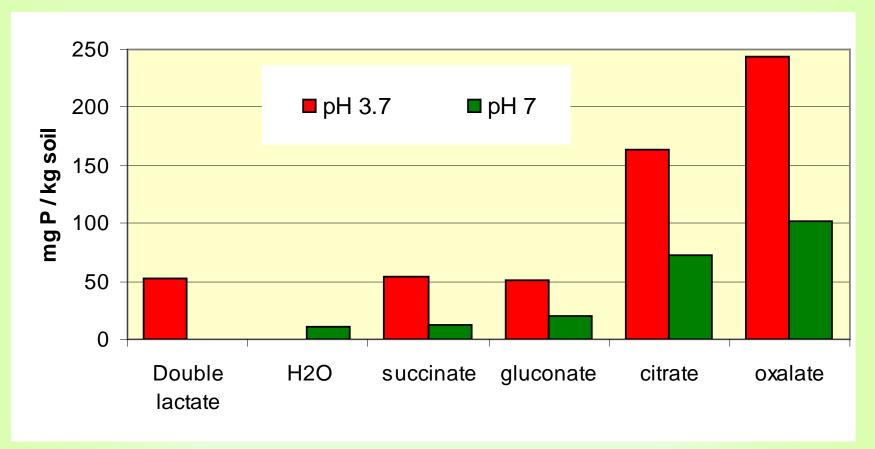


P concentration in soils (mg kg⁻¹)

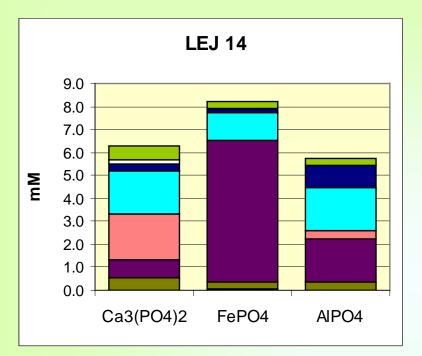


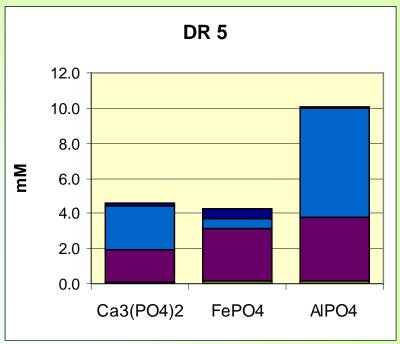
рН	% Fe	% AI
6.0	9.4	6.1
4.8	10.6	5.2
5.2	8.9	5.3
4.3	11.2	5.5
4.0	4.6	5.3
3.7	4.5	5.4
4.1	3.1	4.5
4.0	2.6	4.1
4.9	7.0	3.5
4.1	2.1	3.1
4.5	0.8	1.0
4.8	1.0	1.4
5.0	5.2	4.5
5.5	0.9	1.1
4.7	1.2	1.3
7.4	1.4	1.8

Soil conditions

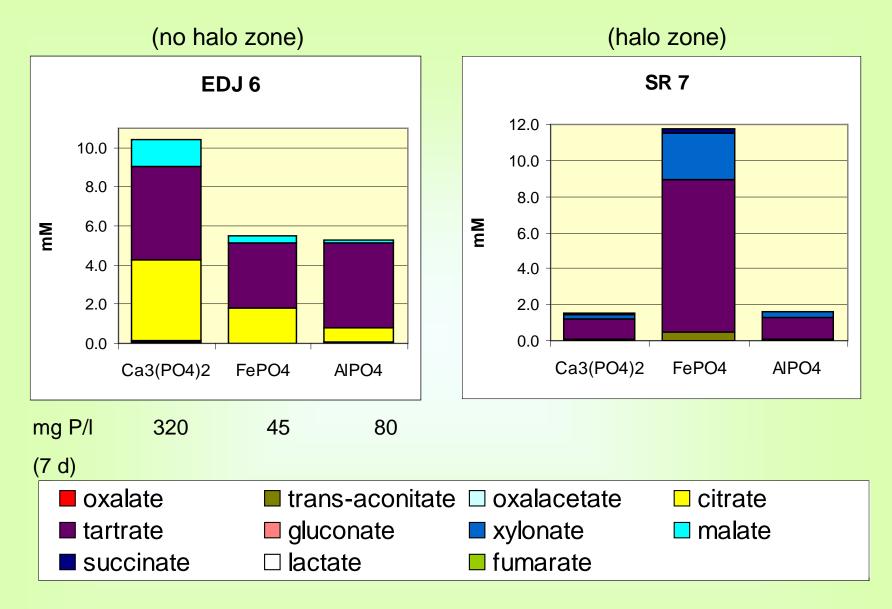


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Strain EDJ 6 = most efficient CaP mobiliser: High citrate production and missing halo zone (precipitation of Ca citrate in agar plate test?)

